

perimental results confirm the presence of N-3-methylbutylacetamide in the venom of the southern yellowjacket and demonstrate alarm pheromone activity of this compound similar to that obtained with venom extracts. This is only the second alarm pheromone of a social wasp identified to date. The alarm pheromone of *Vespa crabro* L. is reported to be 2-methyl-3-butene-2-ol<sup>7</sup>.

N-3-Methylbutylacetamide is reportedly a volatile constituent of tobacco<sup>11</sup>, wines and cheeses<sup>13</sup>. Although the amounts of this chemical in these products are not known, the possibility exists of eliciting an attack from a yellowjacket colony with these or other natural materials containing N-3-methylbutylacetamide. This compound also has been found in the rectal glands of the Queensland fruit fly, *Dacus tryoni* (Froggatt), although its function in gland secretions is unknown<sup>16</sup>.

tion of a proprietary product does not constitute an endorsement or the recommendation for its use by USDA.

- 2 Landolt, P. J., and Heath, R. R., Fla. Ent. 70 (1987) 222.
- 3 Maschwitz, U. W., Nature 204 (1964) 324.
- 4 Maschwitz, U., Dt. ent. Z. 31 (1984) 33.
- 5 Jeanne, R. L., Behav. Ecol. Sociobiol. 9 (1981) 143.
- 6 Jeanne, R. L., Experientia 38 (1982) 229.
- 7 Veith, H. J., Koeniger, N., and Maschwitz, U., Naturwissenschaften 71 (1984) 328.
- 8 Akre, R. D., and MacDonald, J. F., Economic Impact and Control of Social Insects 1986, p. 353.
- 9 Kovats, E., Adv. Chromat. 1 (1965) 229.
- 10 Bellas, T. E., and Fletcher, B. S., J. chem. Ecol. 5 (1979) 795.
- 11 Demole, E., and Berthet, D., Helv. chim. Acta 55 (1972) 1866.
- 12 Schreier, P., Drawert, F., and Junker, A., Z. Lebensmittelunters. Forsch. 157 (1975) 34.
- 13 Kowalewska, D., J. Dairy Sci. 68 (1985) 2165.

1 The authors thank E. Adamak, R. Murphy and F. Takken for technical assistance. This article reports the results of research only. Men-

0014-4754/88/010082-02\$1.50 + 0.20/0  
© Birkhäuser Verlag Basel, 1988

### Goniothalamycin and annonacin: Bioactive acetogenins from *Goniothalamus giganteus* (Annonaceae)

A. Alkofahi, J. K. Rupprecht, D. L. Smith, Ch.-J. Chang and J. L. McLaughlin

Department of Medicinal Chemistry and Pharmacognosy, School of Pharmacy and Pharmacal Sciences, Purdue University, West Lafayette (Indiana 47907, USA), 7 July 1987

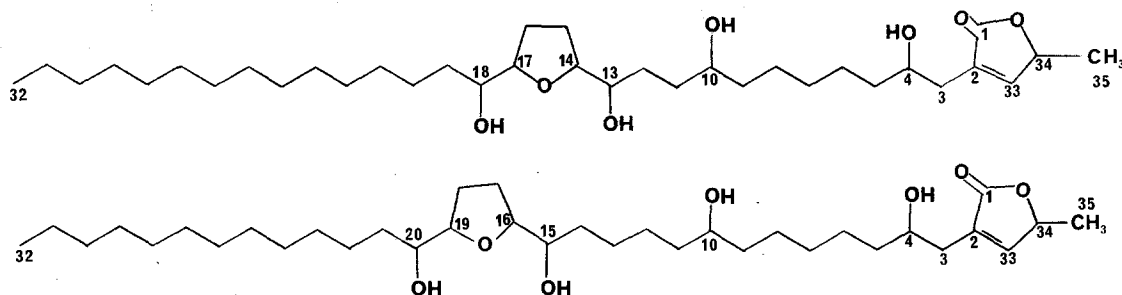
**Summary.** Using brine shrimp lethality for activity-directed fractionation, goniothalamycin (**I**), a new tetrahydroxy-mono-tetrahydrofuran fatty acid  $\gamma$ -lactone (acetogenin), has been isolated from ethanolic extracts of the stem bark of *Goniothalamus giganteus* Hook. f., Thomas (Annonaceae). This novel compound was found to be cytotoxic and insecticidal and inhibited the formation of crown gall tumors on potato discs. Annonacin (**II**), the only other reported mono-tetrahydrofuran acetogenin, was also isolated; the previously reported 9ASK (astrocytoma reversal) activity of **II** was confirmed, and **II** is now also found to be weakly active against 3PS murine leukemia.

**Key words.** Goniothalamycin; annonacin; acetogenins; *Goniothalamus giganteus*; Annonaceae; brine shrimp; insecticidal; cytotoxic; antileukemic activity.

Previous phytochemical studies of the genus *Goniothalamus* have yielded the bioactive compounds altholactone (goniothalenol)<sup>1</sup>, a furano-2-pyrone, and goniothalamycin<sup>1,2</sup>, a styrylpyrone, as well as pinocembrin<sup>1</sup> (5,7-dihydroxyflavone) and a number of 5,6-dihydro-2-pyrone<sup>3</sup>. During our continuing investigation of higher plants as sources of novel biologically active secondary metabolites, we noted that ethanolic extracts of the stem bark of *Goniothalamus giganteus* Hook. f., Thomas (Annonaceae) exhibited significant murine toxicity in the 3PS lymphocytic leukemia system<sup>4</sup>. The fractionation of the ethanolic extract was guided by a convenient bioassay involving brine shrimp lethality<sup>5</sup>. Through multiple solvent partitionings and chromatographic steps, monitoring the fractions with thin-layer chromatography (TLC) and brine shrimp lethality, goniothalamycin (**I**), a novel acetogenin and the recently reported mono-

tetrahydrofuran acetogenin, annonacin<sup>6</sup> (**II**), were isolated and subsequently characterized.

Goniothalamycin (**I**) was found to be cytotoxic in the 9KB (human nasopharyngeal carcinoma, ED<sub>50</sub> < 10<sup>-2</sup>  $\mu$ g/ml) and the 9PS (murine lymphocytic leukemia, ED<sub>50</sub> < 10<sup>-1</sup>  $\mu$ g/ml)<sup>4</sup> systems. This cytotoxic activity paralleled the brine shrimp lethality throughout the fractionation of the stem bark. Goniothalamycin was toxic to the brine shrimp [LC<sub>50</sub> 37 ppm, 95% confidence intervals: 13–295 ppm] and also toxic to blowfly larvae<sup>7</sup> (100% mortality at 1% conc.). In a plant tumor system<sup>8,9</sup>, **I** significantly inhibited the formation of crown gall tumors on potato discs (32% and 28%); this plant tumor system<sup>8,9</sup> has shown a positive correlation with the 3PS murine lymphocytic leukemia system<sup>4</sup> in vivo, indicating that **I** might possess antitumor activity; however, no 3PS activity was observed at



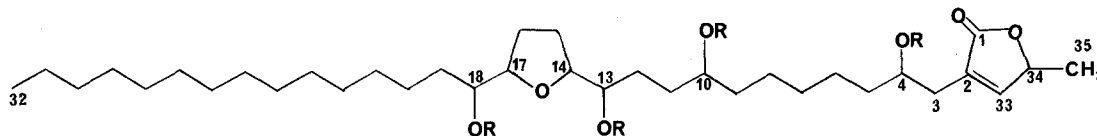


Figure 1. Derivatives of goniiothalamycin analyzed by MS.

I R = H  
 III R = Ac  
 IV R = TMS  
 V R = Ac-d<sub>3</sub>  
 VI R = TMS-d<sub>9</sub>  
 VII R = TMS, 2,33-dihydro

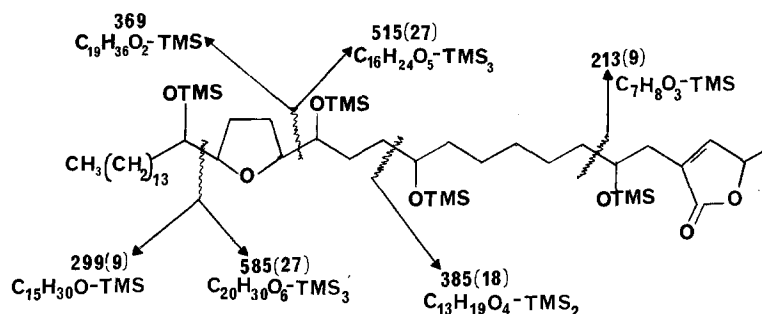


Figure 2. Diagnostic MS fragment ions of IV and VI.

doses up to 2.5 mg/kg; higher doses exhibited murine toxicity. In the 9ASK (astrocytoma reversal) assay<sup>10</sup>, goniiothalamycin was inactive. Annonacin (II) was ten times as active against brine shrimp (LC<sub>50</sub> 3.3 ppm); it also was insecticidal against blowfly larvae (100% mortality at 1% conc.) and mosquito larvae (70% mortality at 10 ppm) and was active in the potato disc crown gall antitumor system (45% inhibition). The potato disc activity of annonacin was correctly indicative of weak 3PS *in vivo* murine antileukemic activity (T/C 124% at 0.95 mg/kg). Also, the previously reported 9ASK activity of annonacin<sup>6</sup> was confirmed (15–30% reversal at 100 µg/ml).

Goniiothalamycin (I) was isolated as a whitish wax (m.p. 86–88 °C, [α]<sub>D</sub> + 1.6°). The high resolution chemical ionization (isobutane) mass spectra (CIMS) gave an MH<sup>+</sup> at *m/z* 597.4732 (calc. 597.4734) corresponding to the molecular formula C<sub>35</sub>H<sub>64</sub>O<sub>7</sub>. The presence of four hydroxyl moieties was suggested by four successive losses of water (*m/z* 18) from the molecular ion in the CIMS. In addition, the IR spectrum contained a broad absorption at 3450 cm<sup>-1</sup>, consistent with the presence of hydroxyl groups. The CIMS of the acetate derivative (III) and the trimethylsilyl derivative (TMS) (IV) demonstrated that I had four exchangeable protons as evidenced by MH<sup>+</sup> ions corresponding to MH<sup>+</sup> + 4Ac and MH<sup>+</sup> + 4TMS, thus, confirming the presence of four hydroxyl groups.

A positive response to Kedde's<sup>11</sup> reagent suggested the presence of an α,β-unsaturated γ-lactone. A prominent IR carbonyl absorption at 1745 cm<sup>-1</sup> and a UV(MeOH) λ<sub>max</sub> at 215 nm (ε = 3.86) supported the presence of this functionality. Expected resonances in the <sup>1</sup>H NMR and signals in <sup>13</sup>C-NMR<sup>6, 12</sup> confirmed the presence of the α,β-unsaturated γ-lactone. Subsequent <sup>1</sup>H-<sup>1</sup>H decoupling experiments showed coupling between the protons on C(3), H(3a) and H(3b), to the single proton at C(4), establishing the presence of an hydroxyl group at C(4) as in asimicin<sup>12</sup> and annonacin<sup>6</sup>.

In addition to the resonances due to the oxygenated carbons of the lactone and the four hydroxylated carbons at δ 74.3 (d), δ 74.1 (d), δ 71.4 (d) and δ 69.8 (d), the <sup>13</sup>C NMR showed two resonances at δ 82.7 (d) and δ 82.4 (d) also due to oxygen

bearing carbons. These <sup>13</sup>C NMR resonances and their corresponding <sup>1</sup>H NMR resonances at δ 3.83 were directly analogous to similar signals in annonacin (II), indicating the presence of a single tetrahydrofuran moiety.

The placement of the hydroxyl groups alpha to the tetrahydrofuran functionality was established by <sup>1</sup>H-<sup>1</sup>H decoupling experiments that linked the two-proton signal at δ 3.81 to one-proton signals at δ 3.43 and δ 3.38. The subsequent downfield shift of these signals to δ 4.79 (m) in the <sup>1</sup>H NMR of the acetate derivative (III) confirmed this assignment.

The single remaining hydroxyl group gave a one-proton signal at δ 3.61 (m) in the <sup>1</sup>H NMR; irradiation of this resonance indicated that this hydroxyl group was isolated along the alkyl chain.

To determine the placement of the tetrahydrofuran ring and the isolated hydroxyl group along the hydrocarbon chain, mass spectral studies were undertaken. The mass spectra of the underivatized goniiothalamycin (I) often gave irreproducible results due to pyrolysis products and thermal rearrangements. This necessitated the synthesis of several derivatives of I (fig. 1), of which the TMS derivative [bis(trimethylsilyl)acetamide in pyridine] (IV) and the perdeuterio-trimethylsilyl derivative (VI) [TMS-d<sub>9</sub>, bis(perdeuterio-trimethylsilyl)acetamide in pyridine] gave the most reproducible mass spectra. Figure 2 shows the key fragments of derivatives IV and VI and their corresponding elemental compositions as determined by exact mass measurements. The ion at *m/z* 213 in the EI spectrum of IV increased by 9 µm in the spectrum of VI and 2 µm in the spectrum of VII, the TMS 2,33-dihydro derivative (10% Pd/C in EtOH), providing further evidence that the C(4) was hydroxylated. The ion at *m/z* 385, which shifted by 18 µm in the spectrum of VI and 2 µm in the spectrum of VII, indicated that a hydroxyl group is located at C(10). The number of carbons between the two rings was established by the ions at *m/z* 515 and 585.

The length of the hydrocarbon chain attached to the tetrahydrofuran ring was indicated by ions at *m/z* 299 and 369 in the mass spectrum of IV and the ion at *m/z* 308 in the mass spectrum of VI. Homologous ions observed in the EI spectrum of III, V, VI, and VII supported these assignments.

Annonacin (**II**) was obtained as a whitish wax with m.p. 57 °C. The molecular formula of annonacin was established by high resolution CIMS, and the IR,  $^{13}\text{C}$  NMR and  $^1\text{H}$  NMR spectra were identical to the published values for annonacin<sup>6</sup>. Co-TLC with an authentic sample of annonacin in five different TLC systems and optical rotations ( $\alpha_D + 1.4^\circ$ ) showed complete homogeneity, indicating that the two isolates were identical.

**Acknowledgments.** This work was supported by grant No. CA 30909 from the National Cancer Institute, NIH. The 470 MHz NMR instrument was available through the Purdue University Biochemical Magnetic Resonance Laboratory which is supported by NIH grant No. RR1077 from the Biotechnology Resources Program of the Division of Research Resources. Special thanks are due to the Purdue Cell Culture Laboratory for cytotoxicity results, Mr Barnard A. Scott at Eli Lilly and Co. (Greenfield) for pesticide testing, Dr John M. Cassady of Purdue University for a sample of annonacin, and Mr Thomas G. McCloud for helpful discussions.

- 1 ElZayat, A. A. E., Ferrigni, N. R., McCloud, T. G., McKenzie, A. T., Byrn, S. R., Cassady, J. M., Chang, C. J., and McLaughlin, J. L., *Tetrahedron Lett.* 26 (1972) 955.
- 2 Jewers, K., Davis, J. B., Dougan, J., Manchanda, A. H., Blunden, G., Kyi, A., and Wetchapinan, S., *Phytochemistry* 11 (1972) 2025.

- 3 Talapatra, S. K., Basa, D., Deb, T., Goswami, S., and Talapatra, B., *Indian J. Chem.* 24B (1985) 29.
- 4 Geran, R. I., Greenberg, N. H., MacDonald, M. M., Schumacher, A. M., and Abbott, B. J., *Cancer Chemother. Rep.* 3 (1972) protocols 16 and 13.
- 5 Meyer, B. N., Ferrigni, N. R., Putnam, J. E., Jacobsen, L. B., Nichols, D. E., and McLaughlin, J. L., *Planta med.* 45 (1982) 31.
- 6 McCloud, T. G., Smith, D. L., Chang, C. J., and Cassidy, J. M., *Experientia*, 43 (1987) 947.
- 7 Insecticidal testing obtained from Barnard A. Scott of Lilly Research Laboratories, Greenfield, IN.
- 8 McLaughlin, J. L., and Ferrigni, N. R., *Proceedings of Symposium on Discovery and Development of Naturally Occurring Antitumor Agents*, NCI/Frederick Cancer Research Facility, Frederick, Maryland, July 27–29, 1983, p. 912.
- 9 Ferrigni, N. R., Putnam, J. E., Anderson, B., Jacobsen, L. B., Nichols, D. E., Moore, D. S., and McLaughlin, J. L., *J. nat. Prod.* 45 (1982) 679.
- 10 Shoemaker, R. H., Abbott, B. J., MacDonald, M. M., Mayo, J. G., Venditti, J. M., and Wolpert DeFilippis, M. K., *Cancer Treatment Rep.* 67 (1983) 1.
- 11 Tschesche, R., Grimmer, G., and Sechafer, F., *Chem. Ber.* 86 (1953) 1235.
- 12 Rupprecht, J. K., Chang, C. J., Cassady, J. M., McLaughlin, J. L., Mikolajczak, K. L., and Weisleder, D., *Heterocycles* 24 (1986) 1197.

0014-4754/88/010083-03\$1.50 + 0.20/0

© Birkhäuser Verlag Basel, 1988

## 1,1-Dimethyl-5,6-dihydroxyindolinium chloride from a deep water marine sponge, *Dercitus* sp.

S. Kohmoto, O. J. McConnell<sup>1,2</sup> and A. Wright

Harbor Branch Oceanographic Institution, Inc./SeaPharm Project, 5600 Old Dixie Highway, Ft. Pierce (Florida 33450, USA), 11 June 1987

**Summary.** 1,1-Dimethyl-5,6-dihydroxyindolinium chloride (**1a**) was identified from a deep water sample of the marine sponge, *Dercitus* sp., and its structure was elucidated by spectral methods.

**Key words.** 1,1-Dimethyl-5,6-dihydroxyindolinium chloride; marine natural products; *Dercitus*; marine sponge.

Numerous nitrogen-containing metabolites have been isolated from marine sponges<sup>3–5</sup>, but only a small percentage of these metabolites contain a quaternary ammonium functionality. In this note, we report the isolation and identification of 1,1-dimethyl-5,6-dihydroxyindolinium chloride (**1a**), a new marine natural product from a deep water sponge, *Dercitus* sp. Gray, 1867<sup>6</sup>. Two tryptophan derivatives, 2'-de-N-methyl-aplysinopsin (**2a**) and 6-bromo-2'-de-N-methyl-aplysinopsin (**2b**), have been reported from a shallow water sample of *Dercitus* sp.<sup>7</sup>

The sponge was collected northwest of Goulding Cay, Bahamas, in August, 1985, at a depth of 215 m using the Harbor Branch Oceanographic Institution's submersible, the Johnson Sea-Link II. Sequential solvent extraction of the fresh frozen sponge (97 g) with EtOAc and MeOH yielded crude extracts (0.15 g and 2.7 g, respectively). From a portion of the MeOH extract (2 g), **1a** (118 mg) was purified with multilayer planetary coil CCC<sup>8</sup> using a solvent system of  $\text{CHCl}_3$ –MeOH– $\text{H}_2\text{O}$  (5/10/6), followed by recrystallization from MeOH– $\text{CHCl}_3$  (m.p. 244 °C).

The molecular formula of **1a** was deduced as  $\text{C}_{10}\text{H}_{14}\text{NO}_2\text{Cl}$  from elemental analysis of the monohydrate of the chloride salt (calculated for  $\text{C}_{10}\text{H}_{16}\text{NO}_3\text{Cl}$ : C, 51.5; H, 6.86; N, 6.00; Cl, 15.0; found: C, 51.58; H, 6.96; N, 6.02; Cl, 15.62) and high resolution FABMS ( $m/z$  of  $\text{C}_{10}\text{H}_{14}\text{NO}_2$ , 180.1021,

40.4 nm). The presence of a 1,2,4,5-tetrasubstituted benzene ring in **1a** was suggested by the  $^1\text{H}$  NMR singlets ( $\delta$  6.81 (H-4) and 7.09 (H-7), the chemical shifts and multiplicities of the  $\text{sp}^2$  carbons (from proton decoupled and DEPT  $^{13}\text{C}$  NMR experiments in  $d_4$ -MeOH:  $\delta$  124.9 (C-3a, s), 112.5 (C-4, d), 149.5 (C-5, s), 147.7 (C-6, s), 104.4 (C-7, d), and 139.5 (C-7a, s), and the relationship of these  $^{13}\text{C}$  NMR doublets with the  $^1\text{H}$  NMR singlets (from a C–H correlation experiment<sup>9</sup>). The presence of two phenolic hydroxyls in **1a** was suggested by the  $^{13}\text{C}$  NMR singlets with chemical shifts of  $\delta$  147.7 and 149.5, IR bands at 3360 and 3140  $\text{cm}^{-1}$ , the absence of a carbonyl band in the IR spectrum, and the formation of a diacetate (**1b**) upon treatment of **1a** with pyridine and acetic anhydride (**1b**:

